

ABOUT JENE

The Journal of Ecology and the Natural Environment (JENE) (ISSN 2006-9847) is published Monthly (one volume per year) by Academic Journals.

Journal of Ecology and the Natural Environment (JENE) provides rapid publication (monthly) of articles in all areas of the subject such as biogeochemical cycles, conservation, paleoecology, plant ecology etc.

The Journal welcomes the submission of manuscripts that meet the general criteria of significance and scientific excellence. Papers will be published shortly after acceptance. All articles published in JENE are peer-reviewed.

Submission of Manuscript

Please read the **Instructions for Authors** before submitting your manuscript. The manuscript files should be given the last name of the first author

Click here to Submit manuscripts online

If you have any difficulty using the online submission system, kindly submit via this email jene@academicjournals.org.

With questions or concerns, please contact the Editorial Office at jene@academicjournals.org.

Editors

Dr. Abd El-Latif Hesham

Genetics Department, Faculty of Agriculture, Assiut University, Assiut 71516, Egypt

Dr. Ahmed Bybordi

East Azarbaijan Research Centre for Agriculture and Natural Resources, Tabriz, Iran

Dr. Sunil Kumar

Natural Resource Ecology Laboratory, Colorado State University 1499 Campus Delivery, A204 NESB, Fort Collins, Colorado-80526, USA

Prof. Gianfranco Rizzo

University of Palermo
Dipartimeno DREAM — Viale delle Scienze - Building
9. 90128
Palermo,
Italy

Dr. Bahman Jabbarian Amiri

Kiel University, Germany, Ökologie-Zentrum der CAU Abt. Hydrologie und Wasserwirtschaft Olhausen Straße, 75 Kiel, Germany

Dr. Bikramjit Sinha

National Institute of Science Technology and Development Studies, Pusa Gate, Dr. KS Krishnan Marg, New Delhi 110012, India

Prof. Gianfranco Rizzo

University of Palermo
Dipartimeno DREAM – Viale delle Scienze - Building
9. 90128
Palermo,
Italy

Associate Editors

Dr. Marko Sabovljevic

Dept. Plant Ecology, Faculty of Biology, University of Belgrade Takovska 43, 11000 Belgrade, Serbia

Dr. Sime-Ngando Télesphore

CNRS LMGE, UMR 6023, Université Blaise Pascal, 63177, Aubière Cedex

France

Dr. Bernd Schierwater

ITZ, Ecology and Evolution, TiHo Hannover Büenteweg 17d, 30559 Hannover, Germany

Dr. Bhattacharyya Pranab

North-East Institute of Science & Technology Medicinal, Aromatic & Economic Plant Division, North-East Institute of Science & Technology, Jorhat-785006, Assam, India

Prof. Marian Petre

University of Pitesti, Faculty of Sciences 1 Targul din Vale Street, Pitesti, 110040, Arges County, Romania.

Prof. R.C. Sihag

CCS Haryana Agricultural University
Department of Zoology & Aquaculture, Hisar-125004,
India

Prof. Kasim Tatic

School of Economics and Business, University of Sarajevo Trg oslobodjenja 1, 71000 SARAJEVO, Bosnia and Herzegovina

Dr. Zuo-Fu Xiang

Central South University of Forestry & Technology, 498 Shaoshan Nanlu, Changsha, Hunan, China.

Editorial Board

Dr. Zuo-Fu Xiang

Central South University of Forestry & Technology, 498 Shaoshan Nanlu, Changsha, Hunan, China.

Dr.Pankaj Sah

Higher College of Technology, Muscat, Department of Applied Sciences, (Applied Biology) Higher College of Technology, Al-Khuwair, PO Box 74, PC 133, Muscat (Sultanate of Oman)

Dr. Arti Prasad

Mohan Lal Sukhadia University, Udaipur,Rajasthan,india. 123,Vidya Nagar,Hiran Magri, Sector-4,Udaipur,Rajasthan, India

Parviz Tarikhi

Mahdasht Satellite Receiving Station (Postal): No. 80, 14th Street, Saadat Abad Avenue, Tehran 1997994313, Iran

Bharath Prithiviraj

Post Doctoral Research Associate Knight Lab, Dept. of Chemistry & Biochemistry University of Colorado at Boulder USA

Dr. Melissa Nursey-Bray

Australian Maritime College, Tasmania, Australia

Parvez Rana

Department of Forestry and Environmental Science Shahjalal University of Science and Technology Bangladesh

Mirza Hasanuzzaman

Faculty of Agriculture, Sher-e-Bangla Agricultural University Sher-e-Bangla Nagar, Dhaka-1207, Bangladesh

Dr. Giri Kattel

Murray Darling Freshwater Research Centre, La Trobe University 471 Benetook Avenue, Mildura, Victoria 3500, Australia

Dr. M. Rufus Kitto

Faculty of Marine Science-Obhur station, King Abdulaziz University, Jeddah 21589, Saudi Arabia

Dr. Özge Zencir

Kemah Vocational Training School, Erzincan University, Kemah, Erzincan, Turkey.

Dr. Sahadev Sharma

Laboratory of Ecology and Systematics, Graduate School of Engineering and Science, University of the Ryukyus, Senbaru 59, Nishihara, Okinawa-903-0213 Japan

Dr. Hasan Kalyoncu

University of Süleyman Demirel, Faculty of Art and Science, Departmant of Biology, 32100 Isparta/Turkey

Hammad Khan

Department of Zoology and Fisheries, University of Agriculture, Faisalaad, Pakistan

Mirza Hasanuzzaman

Faculty of Agriculture, Sher-e-Bangla Agricultural University Sher-e-Bangla Nagar, Dhaka-1207, Bangladesh

Abdurrahman Dundar

Siirt University, Science and Arts Faculty, Department of Biology, 56000, Siirt, Turkey

Meire Cristina Nogueira de Andrade

College of Agronomic Sciences, São Paulo State University, Brazil.

Imran Ahmad Dar

Dept. of Industries and Earth Sciences, The Tamil University, Ocean and Atmospheric Sciences & Technology Cell, (A Unit of Ministry of Earth Sciences, Govt. of India).

S. Jayakumar

Department of Ecology and Environmental Sciences, School of Life Sciences, Pondicherry University, Puducherry - 605 014, India

Umer Farooq

University of Veterinary & Animal Sciences Lahore, Pakistan

Instructions for Author

Electronic submission of manuscripts is strongly encouraged, provided that the text, tables, and figures are included in a single Microsoft Word file (preferably in Arial font).

The **cover letter** should include the corresponding author's full address and telephone/fax numbers and should be in an e-mail message sent to the Editor, with the file, whose name should begin with the first author's surname, as an attachment.

Article Types

Three types of manuscripts may be submitted:

Regular articles: These should describe new and carefully confirmed findings, and experimental procedures should be given in sufficient detail for others to verify the work. The length of a full paper should be the minimum required to describe and interpret the work clearly.

Short Communications: A Short Communication is suitable for recording the results of complete small investigations or giving details of new models or hypotheses, innovative methods, techniques or apparatus. The style of main sections need not conform to that of full-length papers. Short communications are 2 to 4 printed pages (about 6 to 12 manuscript pages) in length.

Reviews: Submissions of reviews and perspectives covering topics of current interest are welcome and encouraged. Reviews should be concise and no longer than 4-6 printed pages (about 12 to 18 manuscript pages). Reviews are also peer-reviewed.

Review Process

All manuscripts are reviewed by an editor and members of the Editorial Board or qualified outside reviewers. Authors cannot nominate reviewers. Only reviewers randomly selected from our database with specialization in the subject area will be contacted to evaluate the manuscripts. The process will be blind review.

Decisions will be made as rapidly as possible, and the journal strives to return reviewers' comments to authors as fast as possible. The editorial board will re-review manuscripts that are accepted pending revision. It is the goal of the AJFS to publish manuscripts within weeks after submission.

Regular articles

All portions of the manuscript must be typed doublespaced and all pages numbered starting from the title page.

The Title should be a brief phrase describing the contents of the paper. The Title Page should include the authors' full names and affiliations, the name of the corresponding author along with phone, fax and E-mail information. Present addresses of authors should appear as a footnote.

The Abstract should be informative and completely self-explanatory, briefly present the topic, state the scope of the experiments, indicate significant data, and point out major findings and conclusions. The Abstract should be 100 to 200 words in length.. Complete sentences, active verbs, and the third person should be used, and the abstract should be written in the past tense. Standard nomenclature should be used and abbreviations should be avoided. No literature should be cited.

Following the abstract, about 3 to 10 key words that will provide indexing references should be listed.

A list of non-standard **Abbreviations** should be added. In general, non-standard abbreviations should be used only when the full term is very long and used often. Each abbreviation should be spelled out and introduced in parentheses the first time it is used in the text. Only recommended SI units should be used. Authors should use the solidus presentation (mg/ml). Standard abbreviations (such as ATP and DNA) need not be defined.

The Introduction should provide a clear statement of the problem, the relevant literature on the subject, and the proposed approach or solution. It should be understandable to colleagues from a broad range of scientific disciplines.

Materials and methods should be complete enough to allow experiments to be reproduced. However, only truly new procedures should be described in detail; previously published procedures should be cited, and important modifications of published procedures should be mentioned briefly. Capitalize trade names and include the manufacturer's name and address. Subheadings should be used. Methods in general use need not be described in detail.

Results should be presented with clarity and precision. The results should be written in the past tense when describing findings in the authors' experiments. Previously published findings should be written in the present tense. Results should be explained, but largely without referring to the literature. Discussion, speculation and detailed interpretation of data should not be included in the Results but should be put into the Discussion section.

The Discussion should interpret the findings in view of the results obtained in this and in past studies on this topic. State the conclusions in a few sentences at the end of the paper. The Results and Discussion sections can include subheadings, and when appropriate, both sections can be combined.

The Acknowledgments of people, grants, funds, etc should be brief.

Tables should be kept to a minimum and be designed to be as simple as possible. Tables are to be typed double-spaced throughout, including headings and footnotes. Each table should be on a separate page, numbered consecutively in Arabic numerals and supplied with a heading and a legend. Tables should be self-explanatory without reference to the text. The details of the methods used in the experiments should preferably be described in the legend instead of in the text. The same data should not be presented in both table and graph form or repeated in the text.

Figure legends should be typed in numerical order on a separate sheet. Graphics should be prepared using applications capable of generating high resolution GIF, TIFF, JPEG or Powerpoint before pasting in the Microsoft Word manuscript file. Tables should be prepared in Microsoft Word. Use Arabic numerals to designate figures and upper case letters for their parts (Figure 1). Begin each legend with a title and include sufficient description so that the figure is understandable without reading the text of the manuscript. Information given in legends should not be repeated in the text.

References: In the text, a reference identified by means of an author's name should be followed by the date of the reference in parentheses. When there are more than two authors, only the first author's name should be mentioned, followed by 'et al'. In the event that an author cited has had two or more works published during the same year, the reference, both in the text and in the reference list, should be identified by a lower case letter like 'a' and 'b' after the date to distinguish the works.

Examples:

Abayomi (2000), Agindotan et al. (2003), (Kelebeni, 1983), (Usman and Smith, 1992), (Chege, 1998;

1987a,b; Tijani, 1993,1995), (Kumasi et al., 2001) References should be listed at the end of the paper in alphabetical order. Articles in preparation or articles submitted for publication, unpublished observations, personal communications, etc. should not be included in the reference list but should only be mentioned in the article text (e.g., A. Kingori, University of Nairobi, Kenya, personal communication). Journal names are abbreviated according to Chemical Abstracts. Authors are fully responsible for the accuracy of the references.

Examples:

Chikere CB, Omoni VT and Chikere BO (2008). Distribution of potential nosocomial pathogens in a hospital environment. Afr. J. Biotechnol. 7: 3535-3539.

Moran GJ, Amii RN, Abrahamian FM, Talan DA (2005). Methicillinresistant Staphylococcus aureus in community-acquired skin infections. Emerg. Infect. Dis. 11: 928-930.

Pitout JDD, Church DL, Gregson DB, Chow BL, McCracken M, Mulvey M, Laupland KB (2007). Molecular epidemiology of CTXM-producing Escherichia coli in the Calgary Health Region: emergence of CTX-M-15-producing isolates. Antimicrob. Agents Chemother. 51: 1281-1286.

Pelczar JR, Harley JP, Klein DA (1993). Microbiology: Concepts and Applications. McGraw-Hill Inc., New York, pp. 591-603.

Short Communications

Short Communications are limited to a maximum of two figures and one table. They should present a complete study that is more limited in scope than is found in full-length papers. The items of manuscript preparation listed above apply to Short Communications with the following differences: (1) Abstracts are limited to 100 words; (2) instead of a separate Materials and Methods section, experimental procedures may be incorporated into Figure Legends and Table footnotes; (3) Results and Discussion should be combined into a single section.

Proofs and Reprints: Electronic proofs will be sent (email attachment) to the corresponding author as a PDF file. Page proofs are considered to be the final version of the manuscript. With the exception of typographical or minor clerical errors, no changes will be made in the manuscript at the proof stage.

Fees and Charges: Authors are required to pay a \$550 handling fee. Publication of an article in the Journal of Ecology and the Natural Environment is not contingent upon the author's ability to pay the charges. Neither is acceptance to pay the handling fee a guarantee that the paper will be accepted for publication. Authors may still request (in advance) that the editorial office waive some of the handling fee under special circumstances

Copyright: © 2014, Academic Journals.

All rights Reserved. In accessing this journal, you agree that you will access the contents for your own personal use but not for any commercial use. Any use and or copies of this Journal in whole or in part must include the customary bibliographic citation, including author attribution, date and article title.

Submission of a manuscript implies: that the work described has not been published before (except in the form of an abstract or as part of a published lecture, or thesis) that it is not under consideration for publication elsewhere; that if and when the manuscript is accepted for publication, the authors agree to automatic transfer of the copyright to the publisher.

Disclaimer of Warranties

In no event shall Academic Journals be liable for any special, incidental, indirect, or consequential damages of any kind arising out of or in connection with the use of the articles or other material derived from the JENE, whether or not advised of the possibility of damage, and on any theory of liability.

This publication is provided "as is" without warranty of any kind, either expressed or implied, including, but not limited to, the implied warranties of merchantability, fitness for a particular purpose, or non-infringement. Descriptions of, or references to, products or publications does not imply endorsement of that product or publication. While every effort is made by Academic Journals to see that no inaccurate or misleading data, opinion or statements appear in this publication, they wish to make it clear that the data and opinions appearing in the articles and advertisements herein are the responsibility of the contributor or advertiser concerned. Academic Journals makes no warranty of any kind, either express or implied, regarding the quality, accuracy, availability, or validity of the data or information in this publication or of any other publication to which it may be linked.

Journal of Ecology and the Natural Environment

Table of Contents: Volume 7 Number 6, June 2015

ARTICLES

Densities, spatial distribution and community structure of plankton of Odot Stream

Imaobong Emmanuel Ekpo, Mandu Asikpo Essien-Ibok and Aniebiet Okon Duncan

The reproductive and feeding ecology of rodents in Sekoru district, Southwest Ethiopia

Tsegaye Gadisa and Kitessa Hundera

academicJournals

Vol. 7(6), pp. 180-187, June, 2015 DOI: 10.5897/JENE2015.0521 Article Number: 162677E53913 ISSN 2006-9847

Copyright © 2015 Author(s) retain the copyright of this article http://www.academicjournals.org/JENE

Journal of Ecology and the Natural Environment

Full Length Research Paper

Densities, spatial distribution and community structure of plankton of Odot Stream

Imaobong Emmanuel Ekpo*, Mandu Asikpo Essien-Ibok and Aniebiet Okon Duncan

Department of Fisheries and Aquatic Environmental Management, University of Uyo, Uyo, Nigeria.

Received 21 April, 2015; Accepted 08 June, 2015

The plankton of Odot Stream, Niger Delta, Nigeria were investigated between June and August, (wet season) and October - December, 2014 (dry season) using standard analytical procedures. The data generated were subjected to descriptive statistics and community structure analysis. A total of 37 species of Phytoplankton belonging to four taxa were identified in ascending order as follows: Xanthophyceae (4.7%), Baccillariophyceae (20.19%), Cyanophyceae (36.34%) and Chlorophyceae (38.71%). Zooplankton belonging to 4 taxa were also identified in ascending order as follows; Cladocera (11%), Copepoda (11%), Larvae (26%), Rotifera (52%). Higher seasonal abundance among the families were recorded in dry season for Chlorophyceae (39.16±14.75), Cyanophyceae (21.83±6.50) and Xanthophyceae (10.66±5.81) while higher wet seasonal abundance were recorded for Baccillariophyceae (22.25±4.59) and Copepoda (0.58±0.34). Chlorophyceae and Rotifera were found as the dominant group in this present study and indicated that Odot stream was highly productive (eutrophic), which could be attributed to high temperature due to the shallowness of the stream and its high exposure to sunlight. Excessive algal bloom should, therefore, be checked to prevent the stream from anoxia and poor water quality and shift in species composition and fish kill.

Key words: Odot Stream, phytoplankton, zooplankton, abundance, Bacillariophyceae, Rotifera

INTRODUCTION

Streams and rivers are very important ecosystems for socio-economic development and sustainability of the environment providing a variety of valuable functions to the environment, national economics and the communities that depend on them (Mbuligwe and Kaseva, 2005; Yillia, 2008). The presence of an aquatic flora or fauna species in an aquatic habitat readily reflects the characteristics of the environment which they exist, showing that one or more ecological variable is within its tolerance range (Dokulil, 2003). The study of plankton

(phytoplankton and zooplankton) is very important because they serve as basis upon which the aquatic ecosystem is supported. The plankton community is a dynamic system that responds quickly to environmental changes; hence they act as indicators of water quality (Wetzel, 2001). Due to several favourable features such as small size, short generation time and a relatively homogenous habitat, plankton organisms are ideal for theoretical and experimental population ecology (Esenowo and Ugwumba, 2010). Several studies have

*Corresponding author. E-mail: imaobongekpo14@yahoo.com.

Author(s) agree that this article remain permanently open access under the terms of the <u>Creative Commons Attribution License 4.0</u> <u>International License</u>

been carried out on various aspects of the aquatic ecosystem in the Niger Delta (Ekpo, 2013; Akpan, 1991; Abowei and George, 2009; Abowei, 2010; Etim et al., 2013; Okiemute and Maduka, 2011). However, no work has been carried out on streams with close proximity to Ibom International Airport one of which is Odot Stream. This study was, therefore, aimed at investigating the plankton composition of Odot Stream as to contribute to the existing knowledge on plankton ecology and distribution and to form S line data for future research.

MATERIALS AND METHODS

Description of study area

Odot stream is located in NsitAtai Local Government Area of Akwa Ibom State, Nigeria. The local government has an area of about 17,000 square metres. It is approximately 50 km from Uyo, the capital of Akwa Ibom State. It is bound in the north by Uruan, in the east by Okobo, in the west by Ibesikpo Asutan and in the south by Uyo L.G.A. with coordinates 4°51'O" N and 8°010 'O" E (Figure 1).

The bottom of the stream is predominantly sandy and muddy while the edges are without mud. The topography of the catchments is sloppy, making water current to be moderately fast from the upstream to the downstream. Activities in the stream include sand dredging, washing of clothes, bathing, and agricultural activities around the stream banks. The climate of the area is typically of tropical rainforests, comprising the dry season (November -March) and wet season (April - October). The dry season is characterized by a prevalence of dry tropical continental winds from the Sahara desert and the wet season by moist tropical maritime winds from the Atlantic Ocean (Ekpo, 2013).

Selection of sampling sites

Water samples were collected from three sampling stations: IkotNyoho Edo - Station 1 (Upstream), IkotMbre- Station 2 (Midstream) and IkotMboho (Downstream) all in Odot, NsitAtai L.G.A. The streams were selected on the basis of the fact that despite its social, economic and ecological role, no published work is available on it.

Sample collection, preservation, identification and analysis

Sampling was done monthly from the three established sites from June-August and October-December, 2014. Since physicochemical and biological features of a lotic ecosystem could vary with time, sampling was done at regular and specific time intervals in the midmorning hours between the hours of 8 am and 12 noon.

Water samples were collected using plankton net with mesh size of 55 mm. The plankton net was horizontally towed for about 3 mins just below the water surface. The samples collected were fixed with approximately 5 ml of 4% formaldehyde solutions in the field immediately after collection and taken to Fisheries Laboratory, University of Uyo for analysis. The preserved samples were allowed to settle first in the sampling bottle, then about two-third of the water sample was decanted into a beaker to concentrate the plankton specimen. 1 ml of the water sample was placed in a Petri-dish and mounted under the microscope for viewing (Egborge, 1973) and appropriate keys (Needham and Needham, 1962; Jeje and Fernando, 1986, 1991; APHA/AWWA/WEF, 2005 and Nwankwo, 2004) were used for identification of the plankton. The total number

of species was recorded as number of cells per ml for phytoplankton; while zooplankton were recorded as number per ml.

Community structure analysis

Three indices were analysed for community structure and include species diversity (Shannon and Weinner, 1949); species equitability or evenness (Pielou 1966) and species richness Margalef (1951).

RESULTS

Analysis of Plankton composition and abundance

A total of thirty seven (37) species of phytoplankton belonging to four taxa were encountered and identified during the study period. The most abundant taxa observed were Chlorophyceae with a percentage composition of 38.79% while the least was Xanthophyceae with percentage composition of 4.75% (Table 1 and Figure 2).

The green algal (Chlorophyceae) were the most important group in terms of diversity with the centric forms recording a high abundance. *Genicularia* sp. dominated with a percentage abundance of 16.38% in the group level and in total number of plankton organism, while the least contributors in terms of number were *Gomphonema sphaeropharum*, *Closterium acerosum*, *Cosmarium quadrum*, *Pediastrum duplex* and *Spirulina* sp., accounting for 0.07% respectively. *Tribonema viridae* was the only species encountered belonging to Xanthophyceae.

In terms of zooplankton, four taxa were identified (Table 2). Rotifera was the most abundant contributing 52% of the composition followed by larvae which contributed 26%, while Cladocera and Copepoda were the least contributing (11% each) (Figure 3).

Seasonal variation among the plankton

The seasonal variation among the plankton was observed and recorded in Table 3. The Baccillariophyceae shows higher seasonal variation in wet season (22.25±4.59) and a lower mean value in dry season (7.66±2.67). Chlorophyceae was observed to be high in dry season (39.16±14.75) and it reduced in wet season (19.33±6.15).

Cyanophyceae showed higher occurrence in dry season (21.83±6.50) and lower occurrence in wet season (20.58±5.44). The family Xanthophyceae showed extremely low occurrence in wet season (3.00±1.48) and higher mean value in dry season (10.66±5.81). Amongst the zooplankton, Rotifera showed high seasonal variation in dry season (3.16±1.07) and no occurrence in wet season.

Copepod showed high value in wet season (0.58±0.43) and low value in dry season (0.166±0.16). Cladocera showed seasonal variation with high mean value in wet

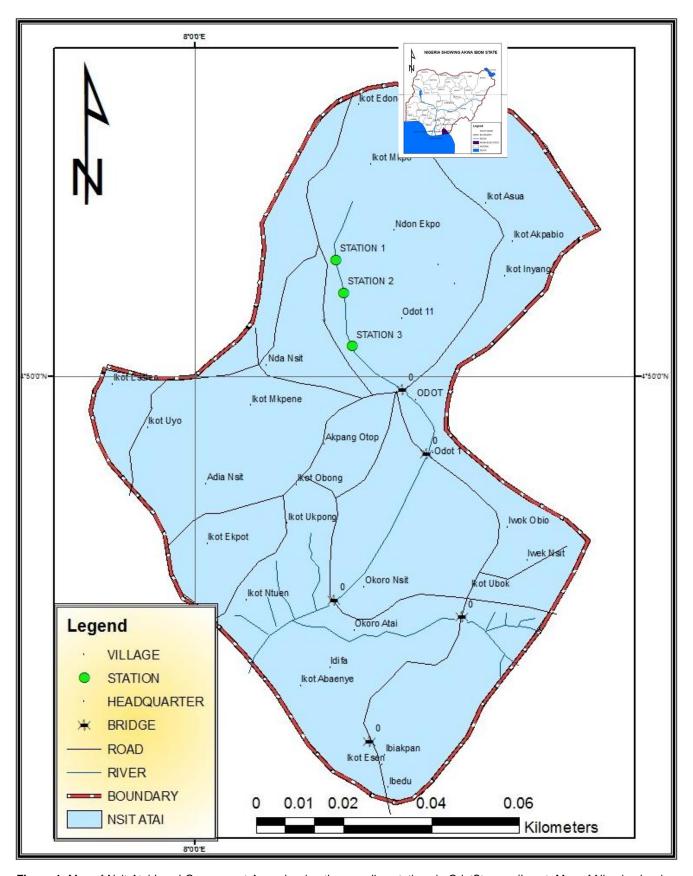


Figure 1. Map of Nsit Atai Local Government Area showing the sampling stations in OdotStream. (Insert: Map of Nigeria showing the location of Akwa Ibom State).

Table 1. Percentage composition and abundance of phytoplankton encountered in Odot Stream.

Plankton diversity	No. of cells/ml	Percentage	e composition %
Baccilariophyceae		_	-
Coscinodiscus radiata	12	0.9	
Cyclotella operculata	20	1.5	
Diatoma sp.	11	0.85	
Epithermia zebra	4	0.31	
Gyrosigma atternuatum	43	3.3	
Melosira listaus	2	0.15	20.19%
Navicula placentula	2	0.15	
Pinnularia major	7	0.54	
Stauroneis sp.	45	3.5	
Tabellaria spp.	22	1.7	
T. fenestra	26	2	
T. fevar asterionelloides	27	2.1	
Thallassiosiren sp.	22	1.7	
Thallassiothrixnitz schioides	11	0.85	
Gomphonema sphoeropharum	1	0.08	
Chlorophyceae			
Ankistrodesmus sp.	5	0.39	
Closterium Cynthia	5	0.39	
Closterium venus	17	1.3	
C. moniliferum	4	0.31	
C. acerosum	1	0.08	
Cosmarum quadrum	1	0.08	38.71%
Dictyosphaerum planctonica	2	0.15	
Dinobaton sp.	171	13.2	
Genicularia sp.	207	15.9	
Gonatozygona culeatum	12	0.92	
Gymnozya moniliferum	26	2	
Microsteria papillifera	2	0.15	
Natrium digitus	2	0.15	
Rediastrum duplex	1	0.08	
Pleurotaenium coronatum	33	2.5	
Cyanophyceae			
Anabaena circinalis	57	4.4	
Oscillatoria planctonica	58	4.5	
O. sancta	173	13.3	36.34%
Phormidium foreolarum	32	2.5	
Phormidium tenue	138	10.6	
Spiruliria sp.	1	0.08	
Xanthophyceae			
Tribonema viridae	60	4.6	4.75%

season (0.50 ± 0.28) and no occurrence in dry season. Larvae were observed to show high value in dry season (0.66 ± 0.33) and low mean value in wet season (0.58 ± 0.28) .

Community structure

The community structure of plankton was computed as shown in Table 4. The taxa with the highest occurrence

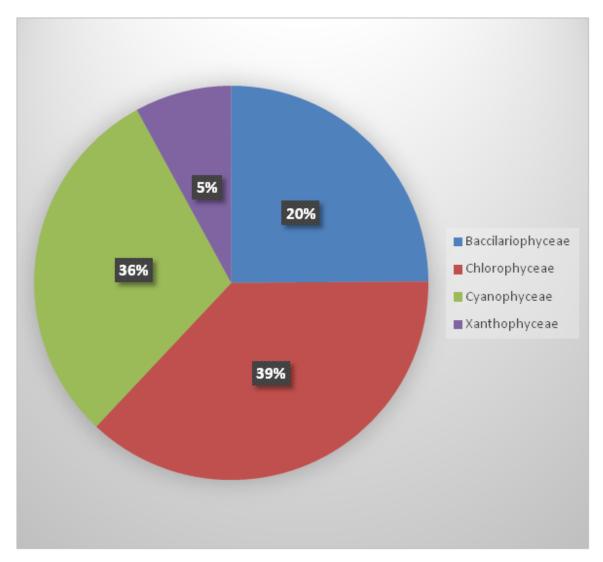


Figure 2. Percentage proportion of the various phytoplankton families in Odot stream.

Table 2. Percentage composition and abundance of zooplankton encountered in Odot Stream.

Zeenlankton tovo	No. of cells/ml	Composit	ion (%)	
Zooplankton taxa	No. of cells/fill	Species	Taxa	
Rotifera				
Asplanchna periodontal	5	0.38		
Philodina sp.	12	0.92	52%	
Trichocera tophoessa	1	0.08		
Cladocera				
Daphnia magnus	2	0.15		
Alonadia phnia	2	0.15	11%	
Copepoda				
Temora longisornis	4	0.31	11%	
Larvae				
Egg larvae	2	0.15	26%	
Nauplii	7	0.54		

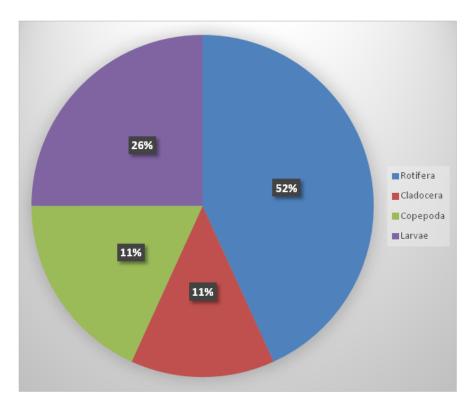


Figure 3. Percentage proportion of the various zooplankton families in Odot stream.

 Table 3. Seasonal variation of plankton encountered in Odot stream.

Plankton	Mean± S. E. (min-max) Wet season	Mean± S. E. (min-max) Dry season				
Phytoplankton						
Dagoillarianhyagaa	22.25±4.59	7.66±2.67				
Baccillariophyceae	(0 - 59)	(2 - 20)				
Chlaraphyagaa	19.33±6.15	39.16±14.75				
Chlorophyceae	(0 - 75)	(4 - 84)				
Cyananhyaaaa	20.58±5.44	21.8±36.50				
Cyanophyceae	(0 - 62)	(2 - 39)				
Variable and because	3.00±1.48	10.66±5.81				
Xanthophyceae	(0 - 16)	(0-36)				
Zooplankton						
Rotifera	0.00±0.00	3.16±1.07				
Rolliera	(0.0 - 0.00)	(0 - 7.00)				
Ola da cara	0.50±0.28	0.00±0.00				
Cladocera	(0 - 0.00)	(0 - 0.00)				
	0.58±0.43	0.166±0.166				
Copepoda	(0.0 - 5.00)	(0.0 - 1.00)				
	0.58±0.28	0.66±0.33				
Larvae	(0.0 - 3.00)	(0.0 - 3.00)				

Table 4. Community structure of plankton.

Donomoton	June		July			August		(October		November			December				
Parameter	1	2	3	1	2	3	1	2	3	1	2	3	1	2	3	Station 1	Station 2	Station 3
Taxa_S	11	6	5	12	12	9	11	4	6	10	10	11	12	9	9	12	9	7
Individuals	112	62	54	150	117	82	74	26	56	26	34	24	142	40	39	106	113	53
Dominance_D	0.38	0.27	0.24	0.17	0.19	0.48	0.18	0.39	0.41	0.25	0.25	0.18	0.24	0.26	0.23	0.17	0.24	0.49
Simpson_1-D	0.61	0.73	0.76	0.82	0.80	0.51	0.81	0.60	0.59	0.75	0.74	0.84	0.76	0.74	0.77	0.82	0.76	0.51
Shannon_H	1.49	1.47	1.49	1.96	1.89	1.22	1.95	1.04	1.19	1.82	1.79	2.03	1.77	1.69	1.83	1.96	1.65	1.12
Evenness_e^H/S	0.40	0.72	0.89	0.59	0.54	0.37	0.63	0.70	0.54	0.62	0.61	0.69	0.49	0.61	0.69	0.59	0.58	0.43
Margalef	2.12	1.21	1.01	2.13	2.31	1.81	2.32	0.92	1.24	2.76	2.55	3.14	2.22	2.16	2.18	2.35	1.69	1.52
Equitability_J	0.62	0.82	0.93	0.79	0.75	0.55	0.81	0.75	0.66	0.79	0.77	0.84	0.72	0.77	0.83	0.79	0.75	0.56

(12) was observed during the months of November in Station 1. December in Station 1 and July in Station 1 and 2. The lowest occurrence in taxa was observed in August, Station 2 being 4. Individual index showed the highest occurrence in July, Station 1 being 150 followed by 142 which occurred in November, Station 1.The lowest individual index was observed in October. Station 3 (24). Dominance index showed high value in December, Station 3 followed by Simpson which showed highest in October, Station 3 (0.84) and lowest in December and July, Station 3 respectively. Shannon- weaver index was observed to be higher in October, Station 3 being 2.03 and lowest in August, Station 2 (1.04). Evenness-e^H/S index was observed with high value in June, Station 3 (0.89) and lowest in June, Station1 (0.40). Equitability index showed higher value in June, Station 3 (0.95) and a lower value in December. Station 3 (0.56).

DISCUSSION

Phytoplankton composition of Odot Stream

The abundance of Chlorophyceae (green algae)

in this present study corresponds with the report of Kemdirim (1990) and Subhashree and Patra, (2013). It shows contrast with those of Essienlbok et al. (2010), Onyema (2010) and Esenowo and Ugwumba (2010) which reported Bacillariophyceae as highest. Among the chlorophyceae, Genicularia sp. was the most abundant species contributing 16.38% of the total plankton species. This was evidenced most in dry season. As observed in the phytoplankton community structure, Dominanice-D index showed high value in December (0.49). Simpson index revealed highest value in December, station 1 and lowest in July, and Station 3. Individual occurrence ranged from 4-12.

Shannon-H index ranged from 1.04 (August Station1) to 2.03 (October, Station 3). Evenness ranged from 0.37 (July, Station 3) to 0.89 (June, Station 3). The abundance of Chlorophyceae is attributed to high water temperature which would cause high photosynthesis and bloom.

Zooplankton

The abundant group among the zooplankton was rotifer. This corresponds with the findings of Ekpo

(2013), and Arimoro et al. (2007). The abundance of rotifer in the stream would be attributed to their ability to undergo vertical migration, which minimizes competition through niche exploitation and food utilization. Also, rotifer richness in the stream probably could be attributed to their pathenogenetic reproductive pattern and short development rates under favorable conditions in most fresh water system (Akin-Oriola, 2003). The author further reported that the rotifers have the shortest life cycle with peak reproductive period of 12 days at 20°C and 5 days at 25°C are normally considered as good indicators of good water quality. Vladimir (1983) suggested that high occurrence of rotifers in any water body indicates an aerobic condition. The result of this study suggests that the rotifer philodinids were more tolerant to environmental stress and that makes it the most occurring organism. This also corresponds with the documentation of Ogbeibu (1998) and Ovie (1997).

Conclusion

Plankton has been used as an indicator to observe and understand changes in the ecosystem.

The high relative abundance of chlorophyceae and rotifers indicate that Odot stream is highly productive (eutrophic), which could be attributed to high temperature due to the shallowness of the stream and its high exposure to sunlight. Excessive algal bloom should, therefore, be checked to prevent the stream from anoxia and poor water quality and shift in species composition and fish kill.

A holistic approach to the management of this stream would be to integrate fish production into its water quality management programme as this would enable the evaluation of the present and future conditions of the stream and its ability to sustain present and future needs.

Conflict of interests

The authors did not declare any conflict of interest.

REFERENCES

- Akin-Oriola GA (2003). Zooplankton associations and environmental factors in Ogunpa and Ona River, Nigeria. Revised Biol. Trop. 51(2):391-398.
- Akpan ER (1991). Seasonal variation in Phytoplankton biomass and pigments in relation to water quality in the Cross River System.Ph.D Thesis, University of Calabar. p. 179.
- APHA, AWWA, WEF (2005). Standard Methods for the examination of water and Wastewater. 21st Edn, American Public Health Association, American Water Works Association and Water Environmental Federation, Washinton, DC, United States.
- Arimoro FO, Ikomi RB, Efemuna E (2007). Macroinvertibrate community patterns and diversity in relation to water quality status of river Ase. Niger Delta, Nigeria. J. Fish Aquat. Sci. 2:337-344. http://dx.doi.org/10.3923/jfas.2007.337.344
- Dokulil MT (2003).Algae as Ecological biomonitors, Elsener Science Ltd. pp. 285-290.
- Egborge ABM (1973). A Preliminary checklist of the phytoplankton of River Oshun. Freshw. Biol. 1(3):569-572. http://dx.doi.org/10.1111/j.1365-2427.1973.tb00078.x
- Ekpo IE (2013). Effect of physico-chemical parameters on zooplankton species and density of a tropical rainforest River in Nigeria Delta, Nigeria using canonical cluster Analysis. Int. J. Engineer. Sci. 2(4):13-21.
- Esenowo IK, Ugwunba AAA (2010). Composition and abundance of Macrobenthes in Majidun River Ikorodu Lagos State. Nigeria. Res. J. Biol. Sci. 5(8): 556-560. http://dx.doi.org/10.3923/rjbsci.2010.556.560
- Essien -Ibok MA, Akpan AW, Udo MT, Chude LA, Umoh IA, Asuquo IE (2010). Seasonality in the Physical and Chemical Characteristics of Mbo River, Akwalbom State, Nigeria. Nig. J. Agric. Food Environ. 6(1-2):60-72.
- Etim EE, Odoh R, Itodo AU, Umoh SD, Lawal U (2013). Water Quality Index for the Assessment of Water Quality from Different Sources in the Niger Delta Region of Nigeria. Front. Sci. 3(3):89-95.
- Jeje CY, Fernando CH (1986). A Practical Guide to the Identification of Nigerian Zooplankton (Cladocera, Copepoda and Rotipfera). Published by KLRI, New Bussa. p. 142.
- Jeje CY, Férnando CH (1991). An illustrated guide to identification of Nigerian freshwater rotifers. Niger. J. Sci. 25:77-95.

- Kemdirim EC (1990). Periodicity and succession of phytoplankton in an upland and lowland impoundments in plateau state(Nigeria)in relative nutrient levels and physical characteristics. J. Aquat. Sci. 5:43-53.
- Mbuligwe SE, kaseva ME (2005). Pollution and self-cleansing of an urban river in a developing country. A case study of Dar Es Salaam, Tanzania Environmental Management, 36(2): 328-342. http://dx.doi.org/10.1007/s00267-003-0068-4
- Needham JG, Needham PR (1962). A Guide to the Study of Freshwater Biology. 2nd Edn, Holden-Day INC., San Franciso, C. A. p 108.
- Ogbeibu AE(1998). Rotifers of a temporary pond in Okonu forest reserve. Niger J. Sci. Environ.1: 117-134.
- Okiemute IE, Maduka K (2011). The impact of industrial effluent discharge on physico- chemical and microbial parameters of Orogodo River, Agbor, Nigeria. Afr. J. Microbiol. Res. 5(22): 3747-3752.
- Onyema IC (2010).Phytoplankton diversity and succession in the lyagbe lagoon, Lagos. Eur. J. Sci. Res. 43(1):61-74.
- Ovie SI (1997).Branchionus species of some inland waters in Nigeria with a note on a new record and zoogeography. Tropical Freshwater Biol. 6:27-39.
- Shannon CE, Weinner W (1949). The mathematical theory of communication. Urban University, Illinois press, Illinois. p. 125.
- Subhashree P, Patra AK (2013). Studies on seasonal variations in phytoplankton diversity of River Mahanadi, cuttack city, Odioha, India. Indian J. Sci. Resour. 4(2):211-217.
- Vladimir SC (1983).Rotifers as indicators of water quality. Hydrobiologia. 100(1).169-201.
- Wetzel RG (2001). Limnology: Lake and River ecosystems, 3rd edition. Academic Press, New York 1006 p.
- Yillia PT (2008). Linking land use to stream pollution, pollutant dynamics and management implications. PhD Dissertation. Vienna University of Technology, Australia.

academicJournals

Vol. 7(6), pp. 188-195, June, 2015 DOI: 10.5897/JENE2015.0516 Article Number: 62B55E753915 ISSN 2006-9847 Copyright © 2015 Author(s) retain the copyright of this article

Journal of Ecology and the Natural Environment

Full Length Research Paper

http://www.academicjournals.org/JENE

The reproductive and feeding ecology of rodents in Sekoru district, Southwest Ethiopia

Tsegaye Gadisa* and Kitessa Hundera

Department of Biology, Jimma University, P.O.Box 378, Jimma, Ethiopia.

Received 30 March, 2015; Accepted 27 April, 2015

A study on reproductive and feeding ecology of rodents was conducted in agricultural areas of some selected kebeles (the smallest administrative unit in Ethiopia) of Sekoru district from February 2014 to December 2014. Rodents were captured by snap trapping using rat traps. During the whole study period, four rodent species, namely, *Rattus rattus, Mastomys natalensis, Arvicanthis dembeensis* and *Lemniscomys barbarus* were identified from a total of 326 rodents captured in 1320 trap nights. The proportion of male and female individuals captured was not different from a 1:1 ratio. Scrotal males and perforate females were captured throughout the study period though reproduction was seasonal. Breeding started in the later part of the rainy season and declined at the beginning of the dry season. The average number of embryo counted per pregnant females was 5.57±1.09, 8.65±1.80, 6.38.±1.70 and 4.00 ± 1.41 for *R. rattus, M. natalensis, L. barbarus* and *A. dembeensis*, respectively. The food items identified during the stomach content analysis were grouped into leaves, seeds, animal matter and unrecognized food item. There was no variation in the type of food item identified among the four rodent species. Some variations were observed in the proportion of the different food items consumed by the four rodent species in different trapping sessions. Awareness creation and the need of rodent pest control in all growth stage of crops and after harvest were recommended.

Key words: Arvicanthis, breeding, feeding, *Lemniscomys*, *Mastomys*, *Rattus*, Sekoru.

INTRODUCTION

Pest animals cause a considerable yield loss throughout the world. Among the crop pests, vertebrate pests especially rodents are responsible for much of the loss and farmers often list rodents as one of the most significant pests to their crops (Singleton et al., 1999; Makundi et al., 2005; Sudarmaji et al., 2003; Tuan et al., 2003). Rodents are regarded as the number one group of mammals in terms of the problems they create in agriculture, horticulture, forestry and public health (Makundi et al., 1999; Brown et al., 1999).

In East Africa, pest rodents cause considerable loss of agricultural crops. In Tanzania, for example, they cause an estimated pre-harvest loss of 15% in maize per annum (Mulungu et al., 2003). Mwanjabe and Leirs (1997) reported the damage to be more than 80% in certain cropping seasons and locations during rodent outbreaks. Taylor (1968) reported 20% damage to maize plantation, 34-100% loss of young wheat in some fields and 34% loss of barely after outbreak of rodents in western Kenya in 1962. Earlier studies in Ethiopia have

*Corresponding author. E-mail: tsegayegad@yahoo.com.

Author(s) agree that this article remain permanently open access under the terms of the <u>Creative Commons Attribution License</u> 4.0International License

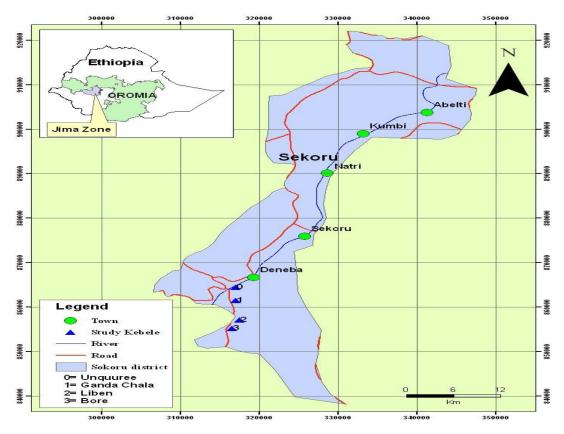


Figure 1. Map of the study area

indicated crop loss of about 20% per annum (Bekele and Leirs, 1997). Later, Bekele et al. (2003) reported an annual yield loss of 26.4% of maize by pest rodents in experimental fields at Zeway (eastern Showa Zone, central Ethiopia) which was even higher than the earlier reports.

Designing and applying effective pest management strategy such as integrated pest management (IPM) can reduce the amount of yield loss caused by pest rodents. Experiences of rodent pest control practices in different countries and research findings to devise appropriate rodent management strategies have showed that successful management of rodent problems depends upon correct identification of the rodent species causing the problem, and obtaining sufficient information on the biology, ecology and behavior of species in the ecological setting where the problem occurs (Tubin and Fall, 2004). Though such ecological studies are essential to devise control strategies of pest species of a particular region, few of such studies were conducted in agricultural areas of Ethiopia (Bekele et al., 1993; Bekele and Leirs, 1997; Lavrenchenko et al., 1998; Gadisa and Bekele, 2006; Bekele et al., 2003; Gebresilassie et al., 2004, 2005, 2006). In spite of serious problems faced by the farmers and many reports on agricultural damages on crops by rodent pests (personal communication with the head of crop protection clinic of the zone), no serious attempts were made in the Jimma zone of southwest Ethiopia to devise rodent control strategy.

MATERIALS AND METHODS

Study area

A study on reproductive and feeding ecology of rodent pests was conducted in some selected kebeles (the smallest administrative units in Ethiopia) of Sekoru district, Jimma Zone, southwest Ethiopia from February 2014 to December 2014 (Figure 1). The altitude of the district ranges from 900 - 2300 m a.s.l. The mean annual temperature of the district is 25°C and the mean annual rain fall is 1500 mm. The wet season is long and ranges from May to October and the dry season ranges from December to April.

Trapping and identification of rodents

Monthly snap trapping was carried out using rat traps for 11 consecutive months (February, 2014 to December, 2014). A total of 30 rat traps were used each day for four consecutive days every month throughout the study period. Traps were set in lines with 10-15 m trap space (Montgomery, 1985; Sutherland, 1996) and peanut butter was used as bait. To increase the trap success, the traps were set along the edges of cultivated fields or in cultivated fields where possible rodent refugia were available or where there were visible runways as rodents use specific paths (Kingdon, 1974;

Trapping	R. I	rattus	L. br	barus	A. den	nbeensis	M. natalensis		
session	Male	Female	Male	Female	Male	Female	Male	Female	
Feb. 14	1	2	2	1	0	0	1	1	
Mar. 14	2	1	1	2	0	0	1	2	
Apr. 13	2	3	2	3	1	2	2	4	
May 14	1	2	3	4	2	2	1	1	
June 14	3	3	2	3	2	1	1	2	
July 14	2	3	3	4	2	1	1	1	
Aug. 14	6	5	7	6	3	2	3	3	
Sep. 14	8	7	13	11	5	3	6	5	
Oct. 14	10	9	17	14	7	5	7	8	
Nov. 14	6	7	7	6	3	4	5	6	
Dec. 14	4	5	6	5	0	1	3	2	
Total	45	47	63	59	25	21	31	35	

Table 1. Species composition and the number of rodents captured by snap trapping at different trapping sessions.

Aplin et al., 2003). After collecting the captured animals, they were identified and standard external measurements were taken. Captured rodents were identified following taxonomic characteristics described by Aplin et al. (2003), Yalden et al. (1996) and Kingdon (1997).

Reproductive conditions

In males, sexual maturity or immaturity was assessed from the conditions of the testes and the scrotal sacs (Aplin et al., 2003; Bekele and Leirs, 1997; Gebresilassie et al., 2006). Captured rodents were considered juveniles if their testes are non-descended or have undeveloped scrotal sacs and sub-adults, if testes were partially descended. Rodents with fully descended testes were considered as adults. In females, the external signs of sexual maturity were assessed from the condition of the vagina and the teats (Aplin et al., 2003, Bekele and Leirs, 1997; Gebresilassie et al., 2006). Females with an open vagina and enlarged teats were considered as adults. A female rodent was considered as currently lactating if the teats are enlarged and the mammary glands are producing milk. Reproductive activities from internal characteristics were assessed after the animals were dissected. A male rodent was considered sexually mature when it possessed large testes, each with prominent blood supply and enlarged epididymis. In a female, features of the uterine horns that can be observed by naked eye or with dissecting microscope was used to determine sexual maturity. Slightly thicker and more elongated uterine horns with a more obvious blood supply but without embryos or placental scars characterize an individual that is entering its first breeding season. Thicker uterine horns with embryo in one or both uterine horns indicate pregnancy (Aplin et al., 2003). The number of embryo in each uterine horn was counted and recorded for pregnant females.

Diet analysis

Out of the snap trapped rodents, 150 were dissected and their stomachs were removed for identifying the diet of rodents in the laboratory following the methods of Leirs (1994). Stomachs were preserved in container containing 10% formalin after identification tag was attached to each stomach until microscopic examination of the stomach samples in the laboratory. In the laboratory, the samples were washed with distilled water to remove fine particles and sieved with sieve of mesh size 0.2 mm for proper identification

and mixed thoroughly. Four slides were prepared for each stomach sample and observed under light microscope to identify the type as well as the proportion of the diet in the sample under study. To quantify the food items of the stomach samples, light microscope with 60x magnification was used. The proportion of different food fragments (leaves of monocots and dicots, seed coats and exoskeleton of arthropods) in the diet was determined by counting the fragments from the entire slide. The presence of food items was recorded in all the prepared slides and was grouped into different types: leaves, seeds, animal matter and unrecognized food item. Chi-square test (χ^2) was used to test the 1:1 ratio of male and female rodents captured at 0.05 levels of significance.

RESULTS

Species composition

From a total of 326 rodents captured in 1320 trap nights, four rodent species were identified: namely, *Rattus rattus, Lemniscomys barbarus, Mastomus natalensis* and *Arvicanthis dembeensis.* Out of the total capture, *Lemniscomys barbarus* comprised the largest proportion of the capture with 123 individuals (37.73%) followed by *R. rattus* with 92 individuals (28.22%). *M. natalensis* and *A. dembeensis* comprised 20.25 and 17.69% of the total capture with 66 and 46 individuals, respectively (Table 1). Among snap trapped species, 164 (50.31%) were males and 162 (49. 69%) were females and this proportion was not significantly different from a 1:1 sex ratio (P = 0.856).

Reproduction

Reproductively active males (scrotal males) and females (perforate females) were captured throughout the whole study period. But, variations were observed in the proportion of scrotal males and perforate females captured in different trapping sessions. For most rodent species identified, the male individuals captured from

Table 2. Reproductive conditions of the four rodent species captured during the study period.

Manth		R. rattus				L. bark	oarus			M. nata	alensis	s	A. dembeensis			
Month	SM	AM	IF	PF	SM	AM	IF	PF	SM	AM	IF	PF	SM	AM	IF	PF
Feb. 14	1	0	0	2	2	0	0	1	1	0	0	1	0	0	0	0
Mar. 14	2	0	0	1	1	0	0	2	1	0	0	2	0	0	0	0
Apr. 14	2	0	0	3	2	0	0	3	2	0	0	4	1	0	0	2
May 14	1	0	0	2	3	0	0	4	1	0	0	1	2	0	0	2
June 14	3	0	0	3	2	0	0	3	1	0	0	2	2	0	0	1
July 14	2	0	0	3	3	0	0	4	1	0	0	1	2	0	0	1
Aug. 14	6	0	0	5	6	1	0	6	3	0	1	2	3	0	0	2
Sep. 14	6	2	2	5	10	3	3	8	4	2	1	4	4	1	0	3
Oct. 14	6	4	3	6	11	6	5	9	4	3	3	5	5	2	1	4
Nov. 14	4	2	2	5	4	3	2	4	3	2	2	4	2	1	1	3
Dec. 14	3	1	1	4	4	2	2	3	2	1	0	2	0	0	0	1
Total	36	9	8	39	48	15	12	47	23	8	7	28	21	4	2	19

SM = Scrotal males, AM = abdominal males, IF = imperforate females, PM = perforate females.

Table 3. Pregnancy rates of adult females of the four rodent species identified at different trapping sessions.

Manth —	R. ra	attus	L. ba	rbarus	M. nat	alensis	A. dembeensis		
Month —	P.F	Pr.F	P.F	Pr.F	P.F	Pr.F	P'F	Pr.F	
Feb. 14	2	0	1	0	1	0	0	0	
Mar. 14	1	0	2	0	2	0	0	0	
Apr. 14	3	0	3	0	3	0	2	0	
May 14	2	0	4	0	4	1	2	0	
June 14	3	1	3	0	3	1	1	0	
July 14	3	1	4	2	4	2	1	0	
Aug. 14	5	3	6	4	6	4	2	1	
Sept. 14	5	3	8	3	8	5	3	1	
Oct. 14	6	3	9	4	9	3	4	2	
Nov. 14	5	2	4	2	4	1	3	1	
Dec. 14	4	1	3	1	3	0	1	0	

P.F = perforate females, Pr.F = pregnant females.

February to August were scrotal and all females were perforate. From September to December, the proportion of scrotal males and perforate females was reduced due to introduction of abdominal males and imperforate females to the population for all the species identified (Table 2). The maximum proportion of abdominal males and imperforate females was recorded in October when more than 30% of the males and females were abdominal and imperforate males and females, respectively.

Among 164 males captured throughout the study period, 128 (78.05%) were scrotal and 133 (82.10%) of 162 females were perforate. In addition, all rodents captured from February to September were scrotal males and perforate females, but, pregnant females were not captured until June for most of the rodent species

identified. The rate of pregnancy was high in August and September for almost all the four species and the rate of pregnancy declined after October (Table 3).

Litter size

Among the 39 perforate females of *R. rattus*, 14 were pregnant. The average number of embryo counted for this species was 5.57±1.09. The number of embryo counted per pregnant female ranged between four and eight. Embryo counts of five and six were more common while that of eight was the least frequent embryo count (Figure 2). Among 47 *L. barbarus* adult females captured during this study, 16 were pregnant. The maximum number of embryo counted for this species was 10 and

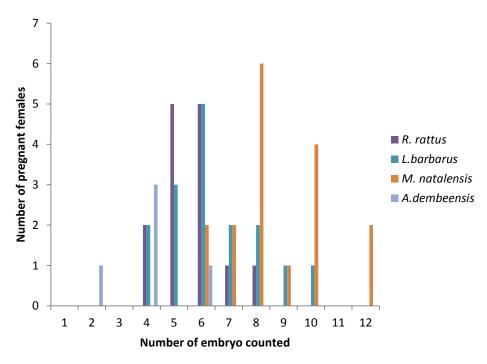


Figure 2. Litter size distributions of the four rodent species identified in the study area.

Canan	Consina	Number of stemashe	Percentage of food items identified							
Season	Species	Number of stomachs	Leaves	Seeds	Animal matter	Unrecognized food				
	R. rattus	23	45.28	27.88	16.75	10.08				
\\/o+	M. natalensis	16	44.95	25.23	18.13	11.63				
Wet	A. dembeensis	23	66.55	18.23	8.88	6,17				
	L. barbarous	32	65.40	18.92	9.20	6.32				
	R. rattus	17	44.96	30.90	14.80	9.52				
D	M. natalensis	14	43.60	30.80	17.00	8.60				
Dry	A. dembeensis	7	61.17	22.43	6.93	9.77				
	L. barbarous	18	60.36	22.88	7.32	9.68				

the minimum count was four. The most frequently counted embryo number was six (Figure 2). The average litter size for the species was 6.38±1.70 For *M. natalensis* and *A. dembeensis*, 17 and 5 pregnant females were captured from 47 and 17 perforate females captured, respectively. The number of embryo counted ranged from 6 to 12 for *M. natalensis* and from two to six for *A. dembeensis* (Figure 2). The average litter size recorded for *M. natalensis* in this study was 8.65±1.80. For *A. dembeensis*, the mean number of embryo counted per pregnant female was 4.00±1.41.

Diet

Stomach samples of 40 R. rattus (20 males and 20

females), 30 A. dembeensis (15 males and 15 females) and 30 M. natalensis (15 males and 15 females) and 50 L. barbarus (25 males and 25 females) were analyzed thought-out the study period. Some variations were observed in the proportion of food items the four different rodent species consumed during dry (November to April) and wet (May to October) seasons (Table 4). For R. rattus, the consumption of leaves was generally high in both seasons but it consumed relatively more leaves in the wet season than the dry seasons. Seed consumption of this species was high in the dry season and the consumption of animal matter was relatively more in the wet season. M. natalensis consumed more leaves in the wet season and its seed consumption was relatively high in the dry season. The consumption of animal matter was more in the wet season. The consumption of leaves by A.

dembeensis and L. barbarous was more as compared to the consumption of leaves by the other two species. Leaves comprised more than 60% of the food items consumed by these species in both seasons.

DISCUSSION

Among the four rodent species identified in the present study area, *M. natalensis* and *A. dembeensis* were recognized as the major pest species in agricultural fields by different researchers in other parts of Ethiopia (Bekele et al., 1993; Bekele and Leirs, 1997; Bekele et al., 2003; Lavrenchenko et al., 1998). The presence of these important pest species in the study area suggests the potential of serious crop damages and losses by them and should be major concerns to the farmers in the area. In addition, all *Mastomys* species in Africa are reported to cause problems in agriculture and public health (Leirs et al., 1996; Fiedler, 1988a, b; Odhiambo and Oguge, 2003). This underlines the need of controlling these species in the study area though their number in the present investigation was low.

The other important species identified in this study are *R. rattus* and *L. barbarous*. In many literatures, *R. rattus* species is mentioned as one of the major pest species that causes a worldwide problem by damaging coconuts and food crops (Fiedler and Fall, 1994; Buckle, 1999). However, *R. rattus*was not reported as a major pest problem in agricultural fields in Ethiopia in reports of earlier research works (Bekele and Leirs, 1997; Bekele et al., 2003).

Many individuals of this species were captured in farmlands with serious damages that were closer to human dwellings. Thus, this species could be a serious problem for farmers who have farmlands closer to their house. *L. barbarus*, though not reported as an important pest species in other parts of Ethiopia, was the dominant pest species in the present study area. Due to its diurnal behaviour, even farmers in the study area were able to see this species climbing the stem of maize and causing serious damage especially during the ripening stage.

Generally, scrotal males and perforate females were captured throughout the study period. But, the proportion of scrotal males and perforate females was lowered in some trapping sessions because of introduction of abdominal males and imperforate females to the population. From the variation in the proportion of scrotal males and perforate females observed, one could say that these species must have started reproduction before the months when reduction in proportion was observed and the new born individuals of both sexes have reached a trappable size at this month and the high proportion of scrotal males perforate females after December suggests that all the new born individuals that entered the population have reached the age of maturity (became adults) and reproduction have ceased to produce more

young individuals of both sexes (Makundi et al., 2007). If breeding has continued, the addition of juveniles into the population continues but this did not happen. In general, it is possible to say that breeding for these four species is restricted to the wet season, especially to the later part of the rainy season and ceases in the dry season. This finding is in agreement with many research results obtained in many rodent pest species in other areas (Bekele and Leirs, 1997; Gadisa and Bekele, 2006; Odhiambo et al., 2005; Leirs, 1994; Lima et al., 2003).

Although the percentage of scrotal males and perforate females was very high in most of the months during the study period, pregnancy for the four snapped species was detected only from May to December. The first pregnant females were captured in May for M. natalensis and May and June for all the four species. Rate of pregnancy has reached peak in July and August and declined towards the dry season and the last pregnant females were captured in December. From this, it is possible to say that breeding commences from the middle of the rainy season and continues up to the early dry season. Such restrictions of breeding to the wet season for many rodent species have been observed by different workers in many different areas (Bekele and Leirs, 1997; Gadisa and Bekele, 2006; Odhiambo et al., 2005; Leirs, 1994; Lima et al., 2003).

The high percentage of scrotal males and perforate females before September and after December can be explained differently. The high percentages obtained before September were because breeding has started in June and the highest percentage of pregnant females was captured in August, and thus, the number of juveniles that could enter the population before September would be very small so that the percentage of scrotal males perforate females in the population would be very high. Although, the high percentage after December would have a different reason. In a month before December, the percentage of scrotal males and perforate females was relatively lower because of the presence of juveniles in the population. The high percentage after December could be because the juveniles that were present in the population have become adults and no juveniles could enter the population as breeding has ceased in the early dry season.

The average number of embryos counted (8.65 ± 1.80) in this study for M. natalensis was lower than the average number of embryos obtained (10.21 ± 4.32) for grass land M. erythroleucus and 12.84 ± 3.26 for maize field) by Bekele and Leirs (1997) at Koka (central Ethiopia). It was also lower than the average number of embryos obtained (11.80 ± 3.40) for the same species by Duplantier et al. (1996) in Senegal. On the other hand, the average number of embryos counted for A. dembeensis in this study (4.00 ± 1.41) is in line with those (4.20 ± 2.20) obtained by Gadisa and Bekele (2006) at Bilalo (southeast Ethiopia) for the same species but lower than

the ones obtained (5.74±2.65) by Bekele and Leirs (1997) for grassland at Koka. But in the same study, Bekele and Leirs (1997) obtained a higher number of embryos (7.42± 2.63) for maize field. The average litter size (5.85±0.34) obtained by Massawe et al. (2007) for *A. neumanni* (a related species) in central Tanzania is also higher than with the average litter of *A. dembeensis* in this study. The variations could be due to differences in environmental conditions such as availability of food.

Most species of rodents appears to be opportunistic feeders (Johnson, 1961). The result of the present study also confirms this. Regardless of the proportional difference, all rodents consumed plant and animal matters. For instance, the stomach contents of *M. natalensis* and *A. Dembeensis* included monocot seeds; monocot leaves and animal matters. However, the stomachs of *A. dembeensis* contained high percentage of grass in addition to other ingredients. This is in line with the finding by Gebresilassie et al. (2004), which have recorded higher percentage of grass and monocot seeds in the stomachs of *A. dembeensis*.

The food items identified in the stomach contents of the four rodent species were the same: all stomach contents consisted of leaves (monocot leaves, dicot leaves and grass), seeds and animal matter (Table 4). One probable reason for this could be less heterogeneity of vegetation in agricultural areas. However, the proportion of the various food items consumed by the different species showed some variations among the species and in the different trapping sessions. Compared the other two species (R. rattus and M. natalensis), the stomach contents of A. dembeensis and L. barbarous consisted of high percentage of leaves. In addition, the percentage of leaves was higher in the wet season as compared to the dry season. This finding is in agreement with the results obtained by Gebresilasie et al. (2004) in Myngus irrigation field, northern Ethiopia, who recorded more grass in the stomach content of A. dembeensis than the stomach content of M. erythroleucus. The percentage of the leaves consumed by the other two species was also higher during the wet season than the dry season. One probable reason for the high consumption of leaves in the wet season could be their relative high abundance than seeds and other plant matters.

Although, starting the latter part of the wet season (the time when crops start maturing) the relative consumption of leaves decreased and the consumption of seeds increased (Table 4). A similar shifting of diet composition was observed by Gebresilasie et al. (2004) by pest rodents of Myngus irrigation field, northern Ethiopia. Similarly, Oguge (1995) found *M. natalensis* to have eaten more grass and dicotyledonous shoots until the development of their preferred cereals and seeds. Leirs (1994) also found *M. natalensis* to consume more grass and arthropods at the beginning of the rainy season and more cereals during the late rainy season and during the early dry season. This shifting of diets of rodents observed

in this study in different seasons indicates that the crops grown in the study area are under the attack of these pest rodents at their different stages of development.

In this study, variation in the consumption of animal matter was observed between the species and between trapping sessions for a species. The percentage of animal matter consumed by *R. rattus* and *M. natalensis* was relatively higher than the animal matter consumed by *A. dembeensis*.

In addition, all the four species consumed more percentage of animal matter during the wet season than the dry season. This variation may be related to the fulfillment of their protein requirement during the seasons of reproduction. The results of this study are comparable with the results of other workers. For example, Leirs (1994) reported that M. natalensis consume more grass and arthropods at the beginning of the rainy season and more cereals during the late rainy season and during the early dry season. Gebresilasie et al. (2004) also reported that reproductively active females consumed more arthropods, monocot shoots and wild herbs than other females. Similarly, Rabiu and Fisher (1989) have revealed that the scrotal males of Arvicanthis sp. have consumed more of monocotyledons and animal matter than the abdominal males.

Most rodents are small, secretive and nocturnal, so are rarely seen even when present in a given area in large numbers (Williams, 1993; Witmer, 2007). They also have a high reproductive potential and short period to reach sexual maturity, thus, can invade large area in a short period of time and cause serious damage (Aplin et al., 2003). Thus, farmers should be aware of the presence of these important pest species and advised to monitor their farmland regularly and carry out rodent control activities in all growth stages of their crops.

Conflict of interests

The authors did not declare any conflict of interest.

ACKNOWLEDGEMENTS

The authors thank Jimma University for funding the research. We are also grateful to the development agents (agricultural extension workers) for their help during data collection and the farmers who allowed as to carry out this study in their farm lands.

REFERENCES

Aplin KP, Brown PR, Jacob J, Krebs CJ, Singleton GR (2003). Field methods for rodent studies in Asia and the Indo-Pacific.Australian center for international agricultural research. Mono. 100. ACIAR, Canberra, Australia.223pp.

Bekele A, Leirs H (1997). Population ecology of rodents of maize fields and grassland in central Ethiopia. Belg. J. Zool. 127:39-48.

- Bekele A, Leirs H, Veshagen H (2003). Composition of rodents and damage estimates on maize farms at Ziway, Ethiopia. In: Singleton GR, Hinds LA, Krebs CJ and Spratt DM (Eds). Rats, mice and people: rodent biology and management. Canberra, Australia: ACIAR. pp. 301-303.
- Bekele A, Capanna E, Corti M, Marcus LF, Schlitter DA (1993). Systematic and geographic variation of Ethiopian Arvicanthis(Rodentia, Muridae). J. Zool. 230:117-134. http://dx.doi.org/10.1111/j.1469-7998.1993.tb02676.x
- Brown PR, Hung NQ, Hung NM, van Wansveen M (1999). Population ecology and management of rodent pests in the Mekong River Delta, Vietnam. In: Singleton GR, Hinds LA, Leirs H, and Zhang Z (Eds). Ecologically-based management of rodent pests. Canberra, Australia: ACIAR. pp. 319-337.
- Buckle AP (1999). Rodenticides- Their role in rodent pest management in tropical agriculture.In: Singleton GR, Hinds LA, Leirs H, and Zhang Z (Eds). Ecologically-based management of rodent pests. Canberra, Australia: ACIAR. pp. 163-177.
- Fiedler LA (1988a). Rodent problems in Africa.In:Prakash J (eds). Rodent pest management.Boca Raton, CRC Press Inc. pp. 35-65.
- Fiedler LA (1988b). Rodent pest problems and management in eastern Africa. FAO Plant Protect. Bull. 36:125-135.
- Fiedler LA, Fall MW (1994). Rodent control in practice: Tropical field crops. In: Buckle AP and Smith RR eds. Rodent pest and their control. CAB International, Wallingford, Oxon, UK. pp. 313-338.
- Gadisa T, Bekele A (2006). Population dynamics of agricultural rodent pest of the Bilalo area, Arsi Ethiopia. Ethiop. J. Biol. Sci. 5(1):63-74.
- Gebresilassie W, Bekele A, Belay G, Balakrishnan M (2005). Population structure of rodents in Maynugus irrigation field, northern Ethiopia. Int. J. Ecol. Environ. Sci. 31:337-342.
- Gebresilassie W, Bekele A, Belay G, Balakrishnan M(2004). Microhabitat choice and diet of rodents in Maynugus irrigation field, northern. Ethiopia. Afr. J. Ecol. 42:315-321. http://dx.doi.org/10.1111/j.1365-2028.2004.00530.x
- Gebresilassie W, Bekele A, Belay G, Balakrishnan M (2006). Home range and reproduction of rodents in Maynugus irrigation field, northern Ethiopia. SINET: Ethiop. J. Sci. 29:57-62.
- Kingdon J (1974). East African Mammals: An Atlas of Evolution in Africa II (b) Rodents and Hares. Academic Press, London.
- Kingdon J (1997). Field Guide to African Mammals. Harcourt Brace and Company Publishers, London.
- Lavrenchenko LA, Likhnova OP, Baskevich MI, Bekele A (1998). Systematics and distribution of Mastomys(Muridae, Rodentia) from Ethiopia, with a description of a new species. Z. Saugetierk. 63:37-51.
- Leirs H (1994). Population Ecology of Mastomysnatalensis (Smith, 1834): Implication for rodent control in Africa. Agricultural Edition. Belgian Administration for Development Cooperation.
- Leirs H, Verhagen, Verheyen R, Mwanjabe WP, Mbise T (1996). Forecasting rodent outbreaks in Africa: an ecological basis for Mastomys control in Tanzania. J. Appl. Ecol. 33:937-943. http://dx.doi.org/10.2307/2404675
- Makundi RH, Massawe AW and Mulugu LS (2007).Reproduction and population dynamics of Mastomysnatalensis Smith, 1834 in agricultural landscape in the western Usambra Mountains, Tanzania. Integr. Zool. 2:233-236. http://dx.doi.org/10.1111/j.1749-4877.2007.00063.x
- Makundi RH, Bekele A, Leirs H, Massawe AW, Rwamugira W, Mulungu LS (2005). Farmers' perceptions of rodents as crop pests: knowledge, attitudes and practices in rat management in Tanzania and Ethiopia. Belg. J. Zool. 135(Supplement):153-157.
- Makundi RH, Oguge NO, Mwanjabe PS (1999). Rodent pest management in East Africa: anecological approach. In: Singleton GR, Hinds LA, Leirs H, and Zhang Z (Eds). Ecologically-based management of rodent pests. Canberra, Australia: ACIAR. pp. 460-76.
- Massawe AW, Mrsso FP, Makundi RH, Mulugu LS (2007).Breeding patterns of Arvicanthisneumani in central Tanzania. Afr. J. Ecol. 46: 320-324. http://dx.doi.org/10.1111/j.1365-2028.2007.00837.x
- Montgomery WL (1985). The effects of recapture and the estimation of Apodedumus spp. J. Zool. London. 206:267-269. http://dx.doi.org/10.1111/j.1469-7998.1985.tb05651.x

- Mulungu LS, Makundi RH, Leirs H (2003). The rodent density-damage function in maize fields at an early growth stage. In: Singleton GR, Hinds LA, Krebs CJ, and Spratt DM (Eds). Rats, mice and people: rodent biology and management. Canberra, Australia: ACIAR. pp. 301-303.
- Mwanjabe PS, Leirs H(1997). An early warning system for IPM-based rodent control in smallholder farming systems in Tanzania. Belg. J. Zool. 127:49-58.
- Odhiambo CO, Oguge NO (2003). Patterns of rodent pest distribution in maize cropping system in the Kenyan Rift Valley. In: SIngleton GR, Hinds LAKrebs CJ and Spratt DM (eds). Rats, Mice and People:Rodent Biology and Management. ACIAR Mono. 96: 217-219.
- Odhiambo R, Makundi R, Leirs Hand Verhagen R (2005).Community structure and seasonal abundance of rodents of maize fields in southwestern Tanzania. Belg. J. Zool. 135 (Supplement): 113-118.
- Oguge ON (1995). Diet, seasonal abundance and microhabitats of Praomys (Mastomys) natalensis (Rodentia; Muridae) and other small rodents in a Kenyan sub-humid grass land community. Afr. J. Ecol. 33:211-223. http://dx.doi.org/10.1111/j.1365-2028.1995.tb00798.x
- Rabiu S, Fisher M (1989).The breeding season and diet of Arvicanthis in northern Nigeria. Trop. Zool. 5:375-386. http://dx.doi.org/10.1017/s0266467400003837
- Singleton GR, Sudarmaji R, Tan TQ, Hung N (1999). Physical control of rats in developing countries. In: Singleton GR, Hinds LA, Leirs H, and Zhang Z (Eds). Ecologically-based management of rodent pests. Canberra, Australia: ACIAR. pp. 178-198.
- Sudarmaji R, Singleton N,Herawati A, Djatherthi A (2003). Farmers' perceptions and practices in rat management in west Java, Indonesia. In:Singleton GR, Hinds LA, Krebs CJand Spratt DM (eds). Rats Mice and People Rodent Biology and Management. ACIAR Monograph 96:389-394.
- Sutherland HM (1996). Ecological census techniques. Cambridge University Press.
- Taylor KD (1968). An outbreak of rats in agricultural areas of Kenya in 1962. East Afr. Agric. For. J. 34:66-77.
- Tuan NP, Williams SJ, Brown P, Singleton GR, Jan JQ, Hue DJ, Ha PT, Hoa P (2003). Farmers' perceptions and practices in rat management in vinhphuc province, northern Vietnam. In: Singleton G.R, LA Hinds, CJ Krebs and DM Spratt (eds). Rats, Mice and People: Rodent Biologyand Management. ACIAR Monograph 96: 399-402
- Tubin ME, Fall MW (2004). Pest control: Rodents, in agricultural societies, from Encyclopedia of Life Support Systems (EOLSS), developed under the Auspices of the UNESCO, Eolss Publishers, Oxford, UK.
- Witmer G (2007). The ecology of vertebrate pests and integrated pest management (IPM). In: M Kogan and P.Jepson (eds.), Perspectives in ecological theory and integrated pest management. Cambridge University Press, Cambridge, UK. pp. 393-410. http://dx.doi.org/10.1017/CBO9780511752353.013
- Yalden DW, Largen MJ, Korck D, Hilman JC (1996). Catalogue of the mammals of Ethiopia and Eritrea. 7. revised checklist, zoogeography and conservation. Trop. Zool. 9:73-164. http://dx.doi.org/10.1080/03946975.1996.10539304

